

foodproof®

Listeria plus *L. monocytogenes* Detection LyoKit Ready Reference Guide

Revision A, November 2023

Product No. KIT230129 (LP), KIT230130 (RP), KIT230369 (DP)

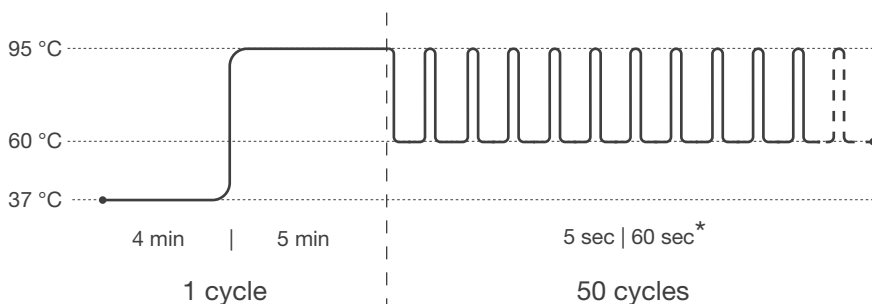
PCR kit for the qualitative detection of the *Listeria sensu stricto* species (*L. monocytogenes*, *L. seeligeri*, *L. ivanovii*, *L. welshimeri*, *L. innocua* and *L. marthii*) including the simultaneous identification of *Listeria monocytogenes*.

Before starting, it is strongly recommended to read the entire product manual available on our website.

PROGRAM SETUP

Program your real-time PCR instrument before setting up the PCR reactions. Select the following channels:

- ▶ FAM (*Listeria monocytogenes*), HEX (*Listeria sensu stricto*) and ROX (Internal Control).



Pre-incubation: 1 cycle

Step 1: 37 °C for 4 min

Step 2: 95 °C for 5 min

Amplification: 50 cycles

Step 1 : 95 °C for 5 sec

Step 2*: 60 °C for 60 sec

* Fluorescence detection

For some real-time PCR instruments the probe quencher as well as the usage of a passive reference dye has to be specified. This kit contains probes with a non-fluorescent "dark" quencher and no passive reference dye.

For the AriaMx real-time PCR instrument, please use transparent low profile tubes (KIT230369, DP).

For the Dualo 32® R2 real-time PCR instrument, please open the software, click on 'New', and select the respective template file. Template files can be added by clicking on 'Add' in the 'Select template file' window.

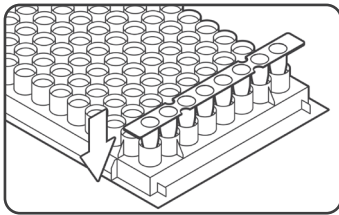
DATA INTERPRETATION

Verify results of positive (Control Template) and negative controls (H₂O), before interpreting sample results. Always compare samples to positive and negative control. Review data from each channel and interpret results as described in the table.

FAM	HEX	ROX	Result Interpretation
+	+	+ or -	Positive for <i>L. monocytogenes</i>
-	+	+ or -	Positive for <i>Listeria sensu stricto</i>
-	-	+	Negative for <i>L. monocytogenes</i> , <i>Listeria sensu stricto</i>
-	-	-	Invalid

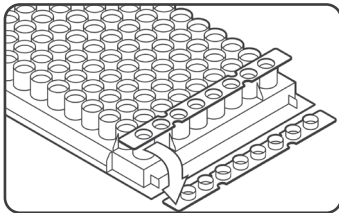
PREPARATION OF THE PCR MIX

Take appropriate precautions to prevent contamination, e.g., by using filter tips and wearing gloves.



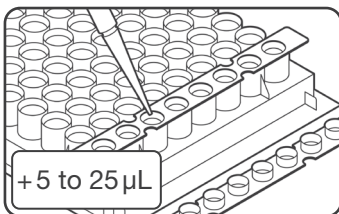
1. PLACE STRIPS IN RACK

Take needed number of PCR tube strips out of aluminum bag. Important: close bag tightly afterwards. Place strips in a suitable PCR tube rack. If needed, gently tap the tubes to move the lyophilized pellets to the bottom of all tubes.



2. DECAP

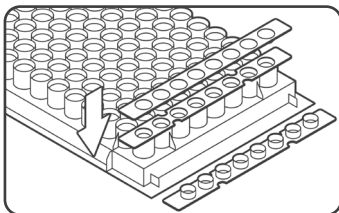
Carefully open strips immediately before filling and discard caps. Do not leave open longer than necessary.



3. ADD SAMPLES AND CONTROLS

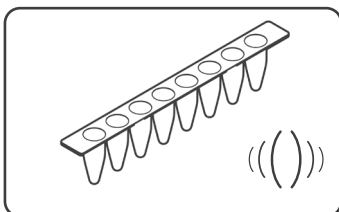
Pipette 5 to 25 µL of samples, 25 µL negative control (colorless cap) or Control Template (purple cap) into respective wells.

Varying sample volumes are due to different enrichment broths and DNA extraction procedures. If using less volume, add PCR-grade H₂O to reach a total volume of 25 µL.



4. SEAL

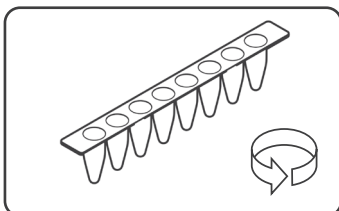
Carefully seal the tubes with the provided 8-cap strips.



5. MIX

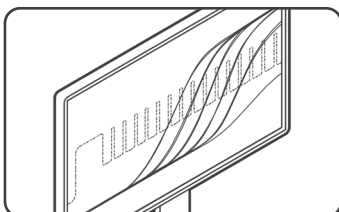
Resuspend pellet after sealing by mixing thoroughly.

Alternatively, resuspend pellet by pipetting up and down multiple times in Step 3.



6. CENTRIFUGE

Briefly spin strips, e.g., 5 seconds at 500 - 1,000 x g, in a suitable centrifuge.



7. START REAL-TIME PCR RUN

Cycle samples as described above.

Place tubes in a vertical, balanced order into the cycler, e.g., two strips can be placed in the first and last column.