

User Guide

Kilo Base (S3) Cartridge Kit (C105106/C105206/C105806)

A. Specifications

Specification	Description
DNA Sizing Range	10-60,000 bp
DNA Detection Range	10-165,000 bp
L.O.D	0.1 ng/μL
Best Resolution*	10-50 bp
Sample Number	200 runs
Shelf Life	6 months

* Best resolution is determined by the 15-622 DNA Size Marker (C109200).

B. Kit Components and Storage Conditions

Item	Storage Condition
Kilo Base Cartridge (C105106/C105206/C105806)	4-30°C (Do Not Freeze)
20-1,000 bp Alignment Marker (C109100-100A, 100 μL)	Short-Term (≤ 3 months): 4-30°C Long-Term (> 3 months): -20°C
Separation Buffer (C104406, 50 mL/C104403, 250 mL)	4-30°C
Dilution Buffer (C104405, 15 mL/C104402, 50 mL)	4-30°C
Mineral Oil (C104404, 8 mL/C104401, 25 mL)	4-30°C

- ⚠ Please always store cartridges in a light-proof bag, and then store in the cartridge box after analysis.
- ⚠ To maintain marker stability, avoid repeated freeze-thaw cycles.

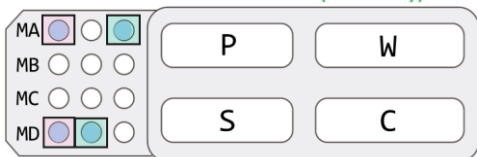
C. Cartridge Unpacking Preparation

The new cartridge must undergo HV check and calibration. Please follow the instructions provided in the unpacking guide and calibrate using C109100 Alignment Marker.

- ⚠ CAUTION: To prevent cartridge lockout after 10 failed calibration attempts, please contact your local distributor or BiOptic if the issue persists after following the troubleshooting steps in the cartridge unpacking guide.

D. Buffer, Marker, and Sample Preparation

MA-1: 20-1k & MA-3: C109700 (Necessary)

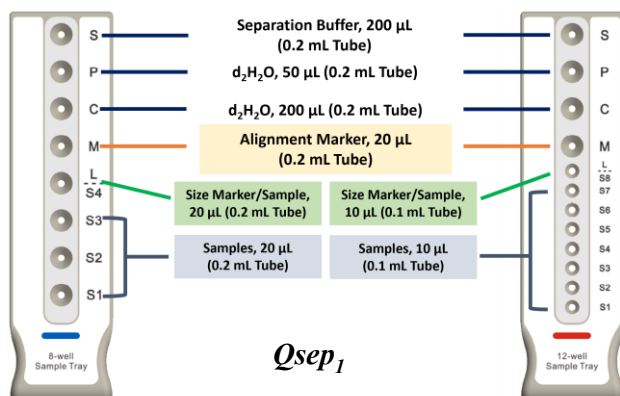


Qsep₁₀₀

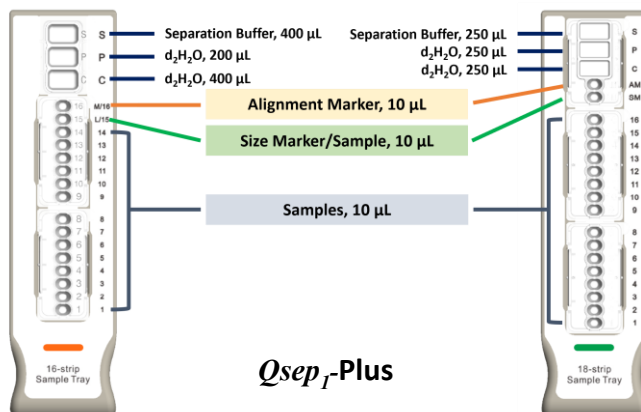
Buffer:
S: Separation Buffer
P/W/C: d₂H₂O
*Add to the groove

MD-1: 20-15k & MD-2: C109400 (Optional)

- ⚠ Hold the holder steady and press the alignment marker tube firmly into the well with your thumb.



Qsep₁



Qsep_{1-Plus}

- ⚠ Ensure the buffer tray is pushed to the end until the color bar aligns with the edge of the holder.

1. Compatible Sample Tubes

	Name	Cat. No.	Volume	Image
Qsep ₁₀₀ 8-well Sample Tray	Micro Vial	C104250	≥ 2 μL	
	0.1 mL PCR Tube	-	≥ 10 μL	
	0.2 mL PCR Tube	-	≥ 20 μL	
*Qsep ₁₀₀ 2.0	0.2 mL PCR Tube	-	≥ 10 μL (≥ 10 μL Mode Only)	
12-well Sample Tray	0.1 mL Strip Tube	C104252	≥ 10 μL	
16-strip Sample Tray	16-strip Sample Tube	C104254	≥ 10 μL	
18-strip Sample Tray	18-strip Sample Tube	C104257	≥ 10 μL	

2. Markers Required for Different Size Ranges

[Recommended]

- For Sample Size Range from 20 bp to 60,000 bp:
 - 20 bp-1,000 bp Alignment Marker (C109100)
 - 500 bp-23,000 bp Size Marker (C109700)
- For Sample Size Range from 20 bp to 15,000 bp:
 - 20 bp-15,000 bp Alignment Marker (C109110)
 - 100-10,000 bp Size Marker (C109400)

- ⓘ Running the C109700 Size Marker is required. Running the C109400 Size Marker is optional. Using a size marker generated under the same running conditions can improve sizing accuracy.

3. Recommended Sample Concentration

Genomic DNA: 2.5-50 ng/μL

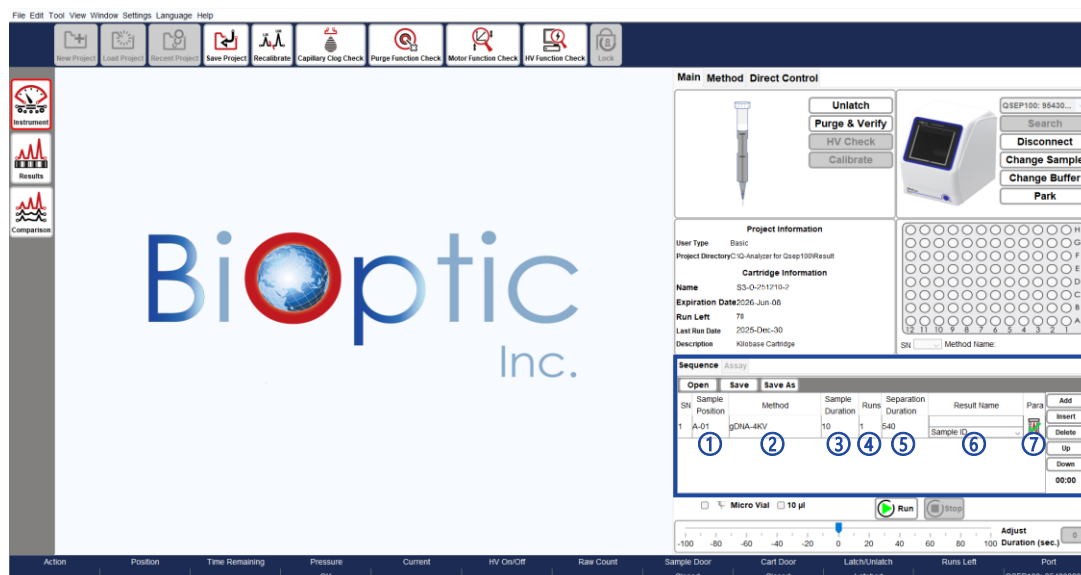
- ⓘ If the sample concentration exceeds 50 ng/μL, dilute the sample 10X using 1X dilution buffer.
- ⓘ If the sample is eluted in water, add dilution buffer to achieve a sample concentration of either 0.2X or 0.1X dilution buffer condition.

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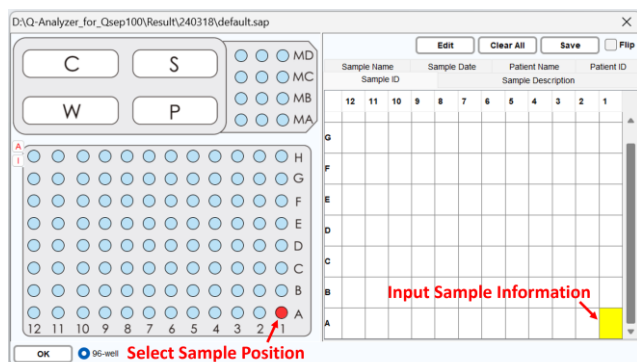
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I. Software Operation Guide

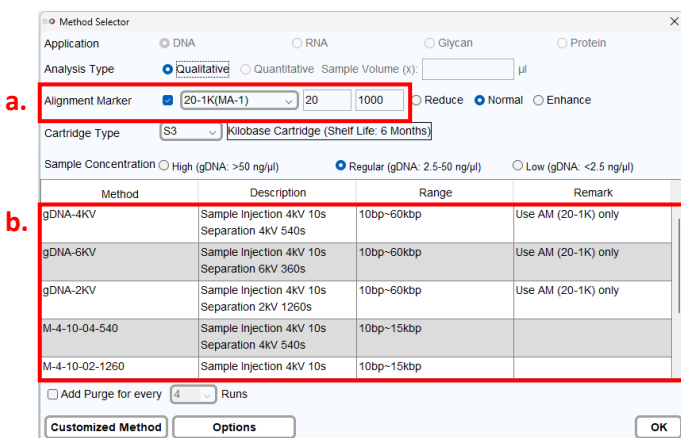
Qsep100



- ① Sample Position: Place the sample and select the corresponding position. Input sample information if necessary.



- ② Method:
 - (a) Select Alignment Marker.
 - (b) Select Analytic Method in the Method Selector.



- Adjust injection conditions based on sample concentration.

Sample Concentration	High (2kV, 10s)	Regular (4kV, 10s)	Low (8kV, 10s)
Genomic DNA	> 50 ng/μL	2.5-50 ng/μL	0.5-2.5 ng/μL

- ③ Sample Duration: Adjust the sample injection time to increase/decrease injection amount.

- Modify injection conditions based on sample concentration.

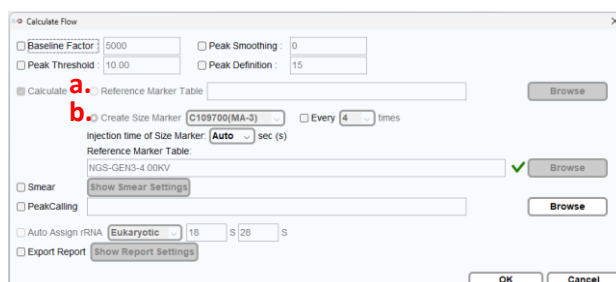
- ④ Runs: Set the repetition time.

- ⑤ Separation Duration: Adjust the duration to extend/reduce the separation time.

(Optional)

- ⑥ Result Name: Input the result name for the result file.

- ⑦ Para: Choose between (a) Reference Marker Table and (b) Create Size Marker for calculation.



- C109700 size marker must be run every 24 hours when using the "gDNA-" methods to obtain sizing results up to 60 kb. If a size marker is created within this timeframe, you have the option to choose "Reference Marker Table" for calculation.

8. Click "Run" to start the sequence analysis.

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Qsep1

- Click "Open Sample Door" and insert the desired sample tray
- Click "Park" to move the sample tray into position.

Qsep1	Qsep1-Plus
8-well Sample Tray 12-well Sample Tray	8-well Sample Tray 12-well Sample Tray 18-strip Sample Tray

① Sample Position:

- Choose Alignment Marker type.
- Position the Alignment Marker at M, place the paired Size Marker at S4 if required, and input the sample information if necessary.

- Adjust injection conditions based on sample concentration.

Sample Concentration	High (2kV, 10s)	Regular (4kV, 10s)	Low (8kV, 10s)
Genomic DNA	> 50 ng/μL	2.5-50 ng/μL	0.5-2.5 ng/μL

③ Sample Duration: Adjust the sample injection time to increase/decrease injection amount.

- Modify injection conditions based on sample concentration.

④ Runs: Set the repetition time.

⑤ Separation Duration: Adjust the duration to extend/reduce the separation time.

(Optional)

⑥ Result Name: Input the result name for the result file.

⑦ Para: Choose between (a) Reference Marker Table and (b) Create Size Marker for calculation.

- C109700 size marker must be run every 24 hours when using the "gDNA-" methods to obtain sizing results up to 60 kb. If a size marker is created within this timeframe, you have the option to choose "Reference Marker Table" for calculation.

8. Click "Run" to start the sequence analysis.

② Method:

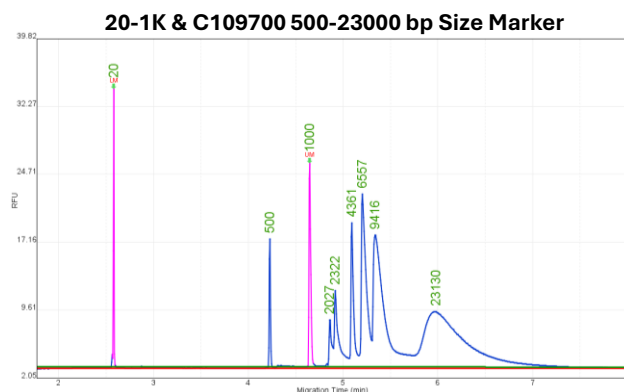
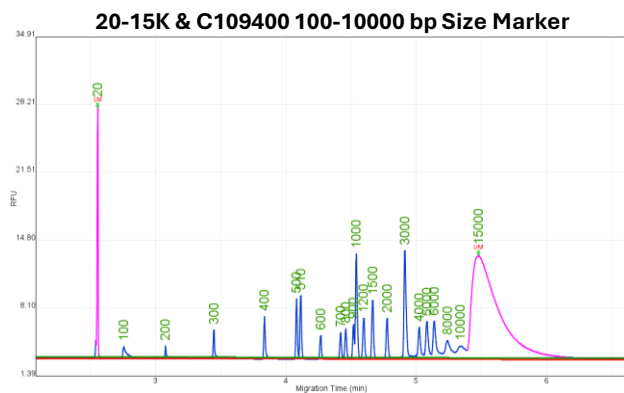
- Check Alignment Marker is ticked.
- Select Analytic Method in the Method Selector.

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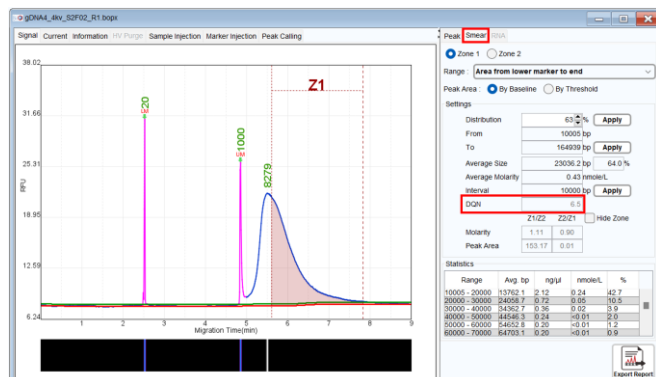
F. Result & Application

Alignment Marker & Size Marker Result



Genomic DNA Validation

Use the smear function to set up a user-defined threshold for DNA Quality Number (DQN) calculation. The DQN, ranging from 1 to 10, can be used as a reference to assess the integrity of genomic DNA.



G. Troubleshooting

Before attempting any troubleshooting, ensure that the system is functioning properly and that all operations are following the instructions.

If encountering unstable current during sample injection or separation steps, which may be caused by residues left in the DNA sample after extraction, consider the following solutions:

1. Dilute the sample using dilution buffer.
2. Centrifuge the sample for a period to allow residues to accumulate at the bottom of the tube.
3. Insert a "T-purge-120" method between several sample runs. For example, insert one run of "T-Purge-120" every 5-10 sample runs.

Sequence		Assay						
Open		Save		Save As				
SN	Sample Position	Method	Sample Duration	Runs	Separation Duration	Result Name	Para	Add
1	A-01.A...	gDNA-2KV	10	1	1260	Test 1 Sample ID		Insert
2		T-Purge-120	0	1	0	Sample ID		Delete
3	B-01.B...	gDNA	10	1	540	Test 2 Sample ID		Up
								Down
								06:30

H. Cartridge Disposal

Please wear gloves before discarding the cartridge.

Gel reservoir



1. Bend the cartridge tip.
2. Open the cap on the gel reservoir and remove the inner cap.
3. Pour the gel into the chemical waste container.
4. Dispose of the cartridge in the trash bin.

Cartridge tip

Contact Information:

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